





OFFICE OF LABORATORY ANIMAL CARE

Operating Guidelines

OG Number:	308	Nonhuman Primate Post-Exposure Protocol for Veterinary Staff	Revision #:	0
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Procedures

1. Patient Care:

- a. The person exposed to the macaque monkey should attend to first aid as soon as it is safe to do so.
- b. Take out a bite/scratch kit
- c. If exposure was a bite/scratch/needlestick, scrub the wound for 15 minutes.
 - i. Remove surgical scrub brush from the kit, apply some water or sterile saline, squeeze a few times to get suds, then scrub the wound site using the soft sponge side of the brush.
- d. If exposure was a splash to mucous membranes (eyes, nose, mouth), patient should go to the nearest eyewash/emergency shower station and flush the area exposed for 15 minutes.
- e. Contact supervisor and veterinarian(s) to notify them of the exposure. If the patient is alone, do this after first aid (scrubbing or flushing wound/exposure site for 15 minutes) has occurred.
- f. Patient takes Herpes B information packet found in the bite/scratch kit to seek medical evaluation:
 - **Regular work hours:** Tang Center Urgent Care (510-642-3188)
 - **Evenings and Weekends:** Alta Bates Emergency Department for physician evaluation.
- g. Patient will tell medical staff at Tang Center or Alta Bates at check-in that there has been a macaque monkey exposure and provide the medical personnel with the Herpes B information packet.

2. Animal Samples

- a. Veterinary staff will take animal samples as soon as possible after human patient care is addressed.
- b. Identify animal(s) or animal cage involved in exposure. If a cage scratch occurred and the animals are pair housed, sample both animals.
- c. Contact PI or other lab personnel to inform them the animal will need to be anesthetized for sample collection.
- d. Anesthetize animal(s) using approved methods.
- e. The following samples should be taken from the animal(s) involved.
 - Blood (5-10 mls) in plain tube or serum separator tube
 - Use only sterile swabs constructed with dacron tips and nylon shafts. Do not use swabs with wooden or metal shafts or cotton tips.
 - Viral swab buccal cavity
 - Viral swab right eye (conjunctiva)
 - Viral swab left eye (conjunctiva)
 - Viral swab genitalia
 - Label with Animal ID number or name, date, and site of swab
 - Place swabs in commercially available viral transport media (VTM) or transport media obtained from B virus lab.
- f. Email the NIH B-Virus Resources Laboratory in Georgia (bvirus@gsu.edu) so that they know to expect samples arriving in the mail.
- g. Do not ship on a Friday or the weekend. Instead, store samples in the refrigerator and ship on the next business day.

3. Sample Preparation (remember to wear gloves)

- a. Use plastic serum separator tubes. Do not ship glass.
- b. Allow blood to clot for 20 minutes.
- c. Spin down blood for 10 minutes
- d. Separate serum into clean plain plastic tube (white top) for transport

4. Storage

- a. Serum
 - Refrigerator storage for up to one week
 - Freezer storage (<-15°C freezer) indefinitely
- b. Swabs
 - Refrigerator storage for up to one week
 - Freezer storage (<-15°C freezer) indefinitely

5. Mailing Samples

- a. Send samples as soon as possible. Do not mail samples on Friday, weekends, or holidays. Send the sample on the next business day.
- b. Fill out the submission form and include it with shipment.
 - This can be found on the Shared drive under Vet-Diagnostic Labs- National B Virus Resource Center or at their website: <http://biotech.gsu.edu/virology>
- c. Pack samples using insulated box and freezer packs.
- d. Send by overnight mail (FedEx) to:

- The National B Virus Resource Laboratory Viral Immunology Center
Georgia State University
161 Jesse Hill Jr. Drive
Atlanta, Georgia 30303

- e. Ship samples as “Dangerous goods”. They **DO NOT** need to be shipped as “Infectious Agent”.
 - Include the following labels:
 - “Biological Substance, Category B” (UN 3373)

- 5. Post Incident Follow-up
 - a. Collect, process, and ship blood samples from the identified animal(s) 14-21 days’ post incident as described above.
 - b. Fax test results received from the Herpes B lab to Occupational Health upon receipt.
 - Fax number: 510-642-6428
 - Cover letter must include:
 - Senders contact information
 - Date of exposure
 - First initial and last name of the human patient

REVISION HISTORY			
REVISION NUMBER	AUTHOR(S)	EFFECTIVE DATE	REVISION(S)